Genetic variants, treatment exposures, and their associations with the development of colorectal cancer as a subsequent malignant neoplasm in long-term survivors of childhood cancer

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BACKGROUND & RATIONALE:

Long-term survivors of childhood cancer face an increased risk of developing subsequent malignant neoplasms (SMNs), with colorectal cancer (CRC) emerging as a significant concern.^{1,2} Compared to the general population, childhood cancer survivors have a 2 to 10.9-fold²⁻⁴ higher risk of developing CRC. CRC as an SMN in this vulnerable population presents a unique challenge due to its potential for early onset and the unknown interplay between genetic predisposition and treatment-related factors.^{3,5-8} While previous research has described genetic risk factors⁹ for CRC in the general population and characterized risk of subsequent CRC in childhood cancer survivors associated with treatment-related exposures (i.e., chemotherapies, radiotherapy),^{2,3,10} there is limited knowledge regarding the role of genetic variants in subsequent CRC development in childhood cancer survivors.

Previous studies through the Childhood Cancer Survivor Study (CCSS)³ and the St. Jude Children's Research Hospital² have demonstrated that the risk of CRC is elevated among childhood cancer survivors who received abdominal/pelvic radiation and alkylating chemotherapy agents, such as procarbazine and cisplatin. In a case-control analysis, the risk of subsequent CRC increased by 70% for each 10-Gray increase in radiation dose to the colon; further, an 8.8-fold increased risk of subsequent CRC was seen with alkylating agent exposure.² Similar findings have been found in cohorts of European childhood cancer survivors.^{4,11,12} On a cellular level, abdominal or pelvic radiation can induce ongoing DNA damage accumulation below the threshold of cell

death in gastrointestinal tissues.¹³⁻¹⁵ Comparably, alkylating agents are genotoxic chemicals that can cause DNA mutations outside of signaling cell death.^{16,17} Both mechanisms can increase the oncogenic potential of cells through genetic alterations.

Genetic predisposition also plays a critical role in SMN development, including CRC. Certain genetic variants associated with hereditary syndromes, such as Lynch syndrome (caused by mutations in MLH1, MSH2, MSH6, and PMS2) and Familial Adenomatous Polyposis (caused by mutations in the APC gene), are known to increase CRC risk. 18-20 However, the role of rare variants in these genes on the risk of developing subsequent CRCs in childhood cancer survivors has not been explored. Across the genes listed in the American College of Medical Genetics and Genomics (ACMG) Secondary Findings, ²¹ in the 60 genes (SJCPG60) known to be associated with autosomal dominant cancer predisposition syndromes, 22 and in the 96 genes inherited in an autosomal dominant or autosomal recessive manner, 22 we have identified 17 genes associated with increased risk of CRC (Table 1). We hypothesize that pathogenic/likely pathogenic variants (P/LP) variants in these 17 CRC-related genes contribute to the development of subsequent CRCs in childhood cancer survivors. particularly for those with early onset CRCs. Early onset CRC is defined as a diagnosis of CRC in individuals at age 50 or younger.²³ Additionally, we hypothesize that genetic risk from pathogenic/likely pathogenic variants in CRC-related genes may be modified by treatment exposures—specifically abdominal/pelvic radiation and alkylating agents (procarbazine or cisplatin)—such that survivors treated with these modalities will exhibit different prevalence and distributions of P/LP variants compared to unexposed survivors across treatment-defined subgroups, reflecting a potential interaction between genetic predisposition and treatment-related DNA damage. Whole-exome sequencing (WES) data from >12.000 survivors of childhood cancer from the CCSS and SJLIFE offers a unique opportunity to preliminarily explore the role of rare P/LP in the risk of subsequent CRCs, as well as their interactions with treatment exposures.

Building on previous work in the CCSS characterizing the risk of subsequent CRCs, our study will address the current knowledge gap described above by leveraging the extensive data available through the CCSS and St. Jude Lifetime Cohort (SJLIFE). Based on an initial review with the CCSS Genetics Working Group, there are approximately 100 survivors with a CRC SMN with WES (31 cases from SJLIFE, 68 cases from CCSS) that can be analyzed for this study. By characterizing the prevalence and distribution of rare P/LP variants in the identified 17 CRC-related genes from the ACMG, SJCPG60, and 96 genes, our study aims to elucidate the interplay between genetic variants and treatment-related exposures related to subsequent CRCs in these survivors. Findings could ultimately play a critical role in the development of targeted interventions, ranging from informing counseling to survivor and survivor families to updating screening guidelines, thus contributing to an improvement in long-term outcomes.

SPECIFIC AIMS:

<u>Aim 1</u>: To characterize the prevalence and distribution of P/LP variants, as listed in Table 1, in survivors diagnosed with a cancer < 21 years of age who subsequently develop CRC and compare results to (i) survivors without a subsequent CRC diagnosis, (ii) individuals in the general population (e.g., SJLIFE community controls), and (iii) an external reference panel (gnomAD) – overall, by treatment exposure, and by age at CRC diagnosis.

<u>Aim 2</u>: To estimate the associations of genetic variants with subsequent CRC in childhood cancer survivors and explore whether primary cancer treatment exposures moderate these associations.

<u>Exploratory Aim 1</u>: To describe the proportion of individuals who received a colonoscopy among childhood cancer survivors who (a) are diagnosed with CRC, and separately (b) report a CRC-related genetic condition.

METHODS/ANALYTIC APPROACH:

Study population (eligibility criteria): Five-year survivors of childhood cancer participating in the CCSS and SJLIFE with WES data. Aim 1 will also utilize SJLIFE community controls.

Dependent/outcome variables:

Development of CRC as a SMN, and age at time of SMN.

Independent/predictor variables:

- Genetic data
 - Whole exome sequencing: We will utilize WES data in the CCSS and SJLIFE cohorts. Currently, WES data of 12,000+ survivors of childhood cancer in the CCSS and SJLIFE are being jointly processed at St. Jude Children's Research Hospital. We expect this process to be complete prior to undertaking this analysis. Once this process is complete, we will create a binary variable (yes/no) indicating carrier status of rare (minor allele frequency, MAF <0.1%)²⁴ P/LP variants in CRC-related cancer predisposition genes to utilize for analysis. We recognize that there is overlap between the CCSS and SJLIFE cohorts; thus, we will keep the identified survivors in SJLIFE and remove the duplicates found in CCSS. We have compiled a list of 17 genes that are associated with increased CRC risk²⁵ across the ACMG, SJCPG60, and 96 genes (genes are listed in Table 1).
 - o In line with previous studies, ^{20,21,26,27} we will analyze variants in cancer predisposition genes with a focus on rare genetic variants. For P/LP variants, the National Center for Biotechnology Information (NCBI) ClinVar²⁸ will be utilized and annotated for these variants. Variants with an aggregate interpretation indicating a consensus in ClinVar will be utilized. In addition, we

will analyze loss-of-function (LOF) and predicted deleterious missense variants. Loss-of-Function Transcript Effect Estimator (LOFTEE; plug-in implemented in the Variant Effect Predictor tool, or VEP²⁹, version 108) will be utilized to annotate LOF variants. If LOF variants are flagged by LOFTEE as "dubious," such as "LOFs affecting poorly conserved exons,³⁰ then they will be excluded. For predicted deleterious missense variants, we will utilize SnpEff³¹ and dbNSFP³² (version 4.1a) for annotations. If more than 90% of annotations predict deleteriousness across the previously mentioned tools/databases, then they will be classified as deleterious.

- Sociodemographic/clinical variables
 - Sex
 - Attained age (i.e., date of birth and date of last contact or death)
 - Vital status (dead/alive)
 - Primary cancer diagnosis
 - o Age at primary cancer diagnosis (i.e., date of primary cancer diagnosis)
 - Reported race/ethnicity
 - Family history of CRC (yes/no)
 - Family history of any cancer (yes/no)
 - Self-reported personal history of colonic polyps (I5 from Follow-up Survey 7 and Follow-up Survey 5; I4 in Follow-up Survey 4; I4 from SJLIFE HOME Survey):
 - Yes" will comprise of self-reported survey answers of:
 - Yes, and the condition is still present
 - Yes, but the condition is no longer present
 - Note: If Yes is marked by the participant, please provide age that participant filled in
 - "No" will comprise of self-reported survey answers of:
 - No
 - Not sure
 - Date of CRC diagnosis and length of time from primary cancer diagnosis to CRC diagnosis
 - o Date of sample collection from participants with genetic data
 - Recurrence of primary cancer and date of diagnosis
 - Other SMN diagnoses and date(s) of diagnosis
 - Specific cancer treatment exposures of interest (delivered within 5 years of primary cancer diagnosis):
 - Abdominal radiotherapy (RT) (yes/no and dose)
 - Pelvic RT (yes/no and dose)
 - Total body irradiation (yes/no and dose)
 - Chemotherapy (yes/no and dose, if available)
 - Alkylating agents

- Yes/no and dose quantified as cyclophosphamideequivalent dose³³ (CED)
- Specific alkylating agents (procarbazine, cisplatin) informing CED (yes/no and dose)
- Self-reported having abdominal or rectal surgery (Item K16 and K17 from Survey Follow up-7; Item J16 and J17 from Survey Follow-up 5; Item J17 and J18 from SJLIFE Home Survey)
 - "Yes" will comprise of self-reported survey answers of:
 - Yes
 - Note: If Yes is marked by participant, please provide age that participant filled in
 - "No" will comprise of self-reported survey answers of:
 - No
 - Not sure
- Self-reported a genetic condition related to CRC (Item W1 from Survey Follow up-7 and Survey Follow-up 5; Item "Hereditary Conditions" from Survey Follow-up 4 and Survey Follow-up 2; Item 4 from Survey Follow-up 1)
 - Survey Follow-up 7 and Survey Follow-up 5:
 - Bloom's syndrome (yes/no/not sure) (Item W1d)
 - Familial adenomatous polyposis (FAP or Gardner syndrome) (yes/no/not sure) (Item W1i)
 - Other (yes/no/not sure and open-ended written response) (Item W1p)
 - In Survey Follow-up 4 and Survey Follow-up 2, if participant circles the following answers under "Hereditary Conditions":
 - Bloom's syndrome
 - Polyposis coli (Gardner syndrome)
 - Survey Follow-up 1:
 - Bloom's syndrome (yes/no/not sure) (Item 4d)
 - Polyposis coli (Gardner syndrome) (yes/no/not sure) (Item 4i)
 - Other (yes/no/not sure and open-ended written response) (Item 4p)
- Self-reported a genetic condition related to CRC (Item P1 from SJLIFE Home Survey)
 - Bloom's syndrome (yes/no/not sure) (Item P1d)
 - Familial adenomatous polyposis (FAP or Gardner syndrome) (yes/no/not sure) (Item P1i)
 - Li-Fraumeni syndrome (p53 gene abnormality) (Item P1r)
 - Other (yes/no/not sure and open-ended written response) (Item P1s)
- Self-reported receipt of colonoscopy (yes/no) (Item C1f from Survey Followup 7, Survey Follow-up 6 Long, and Survey Follow-up 5; Item C4 from Survey Follow-up 4; Item B2 from Survey Follow-up 2)

- "Yes" will comprise of self-reported survey answers of:
 - Less than 1 year ago
 - 1-2 years ago
 - More than 2 years ago but less than 5 years ago
 - 5 or more years
 - I had one but I don't recall when
- "No" will comprise of self-reported survey answers of:
 - Never
 - I don't know if I ever had one
 - Don't know (from Item C4 from Survey Follow-up 4; Item B2 from Survey Follow-up 2)
- Self-reported receipt of colonoscopy (yes/no) (Item N12 from SJLIFE Home Survey)
 - "Yes" will comprise of self-reported survey answers of:
 - Less than 1 year ago
 - 1-2 years ago
 - More than 2 years ago but less than 5 years ago
 - 5 or more years
 - I had one but I don't recall when
 - "No" will comprise of self-reported survey answers of:
 - Never
 - I don't know if I ever had one
- Self-reported body mass index data (Item A1 and A2 from Survey Follow-up 7, Survey Follow-up 6 Long, Survey Follow-up 5, Survey Follow-up 4; Item 7 and 8 in Survey Follow-up 2; Item A3 and A4 from SJLIFE Home Survey)
 - Date of survey collection ("Today's Date")
 - Height
 - Weight
- Self-reported tobacco exposure (yes/no) (Item M7, M9, M10, M11, M13, M14 from Survey Follow-up 7; Item N7, N9, N10, N11, N13, N14 from Survey Follow-up 5 and Survey Follow-up 4; Item L1, L2, L3, L4, L6 from Survey Follow-up 2)
 - "Yes" will comprise of self-reported survey answers of:
 - "Yes" to the question: "Have you smoked at least 100 cigarettes since you last provided us this information"
 - "Yes" to the question: "Do you smoke cigarettes now?"
 - Any number greater than 0 to the question: "On average, how many cigarettes a day do/did you smoke?"
 - Any number greater than 0 to the question: "How many years, in total, have you smoked?"
 - Any positive answer (respondent checks occasionally use or regularly use) to one or more of the items listed (chewing

- tobacco, snuff tobacco, pipes, cigars, or e-cigarettes/vaping) in the question: "In the past year, have you ever used any of these products?"
- Any positive answer (respondent checks less than 1 year, 1-2 years, 3-4 years, 5-10 years, or 11+ years) to the items listed (chewing tobacco, snuff tobacco, pipes, cigars, or e-cigarettes/vaping) in the question: "For any of those that you have used or are currently using, how long how you used it?"
- "No" will comprise of self-reported survey answers of:
 - "No" to the question: "Do you smoke cigarettes now?"

Analysis:

<u>Aim 1</u>: Across the set of 17 CRC-related cancer predisposition genes identified (Table 1), the prevalence and distribution of variants will be tabulated and described among survivors who developed a subsequent CRC and among survivors who did not develop a subsequent CRC. The summary will include variant classes characterization, such as P/LP, LOF, and missense variants, and their frequency. These characteristics will be compared to individuals from the general population via a control sample (the SJLIFE community controls). Since treatment with abdominal/pelvic radiotherapy and specific alkylating agents (procarbazine or cisplatin) has been found to be associated with increased subsequent CRC risk, we will also tabulate distributions of variants for the following subgroups of survivors:

- (i) individuals treated with abdominal/pelvic radiation and no procarbazine or cisplatin
- (ii) individuals treated with procarbazine or cisplatin and no abdominal/pelvic radiation; and
- (iii) individuals not exposed to abdominal/pelvic radiation, procarbazine, or cisplatin Analyses will be repeated restricted to CRC diagnosed prior to age 50 to explore distributions in early-onset CRC.

<u>Aim 2</u>: To evaluate associations between P/LP variants and the development of CRC as an SMN, we will utilize cause-specific Cox regression models with time since diagnosis as the time scale using methods for delayed entry data to reflect that participants enter risks sets at the time a sample for WES was collected. In the primary analysis, all variants across the 17 genes of interest will be grouped, and the presence of any genetic variant will be indicated by a single binary variable (Y/N). In exploratory analysis, if the data permit (large enough numbers of participants with relevant variants), we will repeat the analysis separately for LOF and missense variants. Ideally we would like to analyze associations with variants for each gene separately, but recognize the numbers are likely to be small for such an analysis. We aim to adjust these analyses for sex, age at childhood cancer diagnosis, attained age, family history of CRC, and the genetic ancestry derived from genotype data.

The primary analysis will be repeated adjusting the model for treatment exposures. We will include binary variables of Y/N indicating whether patients received

abdominal/pelvic radiation or alkylating chemotherapies. Our ultimate interest is whether there are interactions between the treatment exposures and the presence of P/LP variant. We can explore this by including interaction terms in our model, but recognize that analyses will be underpowered for formally testing of whether such interactions are present. Further, we will explore whether lifestyle factors available in the data (smoking status and obesity) modify associations, but recognize the limitations of these factors in CCSS survey data. We will adjust for these factors in a simplified manner, based on indicators if participants were ever a smoker or if participants were ever overweight or obese.

Cumulative incidence of CRC will be estimated for survivors stratified by P/LP variant carrier status, accounting for death or lost to follow-up as a competing risk and reflecting the delayed entry data.

Exploratory Aim 1: To obtain the number of childhood cancer survivors who report a CRC-related genetic condition, the total number of individuals who provided a positive response to survey items eliciting a self-reported genetic condition related to CRC will be ascertained via all available CCSS and SJLIFE surveys. A positive response will be counted if the individual reports a CRC-related genetic condition in any of the available CCSS or SJLIFE surveys. Then, the number of positive and negative responses for receipt of a colonoscopy will be determined as per the response groupings from the "self-reported receipt of colonoscopy" variable described above. For participants without CRC, if they ever report having received a colonoscopy in any of the surveys, they will be classified as having received colonoscopy; otherwise, they will be classified as not having received one. For participants with CRC, we will aim to capture responses restricted to surveys completed prior to the CRC diagnosis. To estimate the proportion of individuals who received a colonoscopy among childhood cancer survivors who report a CRC-related genetic condition, the number of individuals who provided a positive response to the "self-reported receipt of colonoscopy" variable will be divided by the total number of individuals who provided a positive response to survey items eliciting a self-reported a genetic condition related to CRC. The same process will be repeated to determine the proportion of individuals who did not receive a colonoscopy among childhood cancer survivors who report a CRC-related genetic condition, except we will utilize the number of individuals who provided a negative response to the "selfreported receipt of colonoscopy" variable as the numerator.

Example tables and figures

Table 1: Complied list of 17 genes associated with increased risk of CRC

HGNC	HGNC approved	Phenotype	Inheritance
symbol*	name		Pattern**
APC	adenomatous	Familial Adenomatous Polyposis	AD
	polyposis coli		

BMPR1A	bone morphogenetic protein receptor, type IA	Hereditary Mixed Polyposis Syndrome	AD
EPCAM	epithelial cell adhesion molecule	Lynch Syndrome	AD
GREM1	gremlin 1, DAN family BMP antagonist	Hereditary mixed polyposis syndrome	AD
MLH1	mutL homolog 1	Lynch Syndrome; Constitutional mismatch repair deficiency	AD; AR
MSH2	mutS homolog 2	Lynch Syndrome; Constitutional mismatch repair deficiency	AD; AR
MSH3	mutS homolog 3	MSH3-associated polyposis	AR
MSH6	mutS homolog 2	Lynch Syndrome; Constitutional mismatch repair deficiency	AD; AR
PMS2	PMS1 homolog 2, mismatch repair system component	Lynch Syndrome; Constitutional mismatch repair deficiency	AD; AR
PTEN	phosphatase and tensin homolog	Cowden Syndrome, increased risk for cancers of the breast, thyroid, endometrium, colorectum, kidney, and skin (i.e. melanoma)	AD
SMAD4	SMAD family member 4	Hereditary Hemorrhagic Telangiectasia; Juvenile Polyposis	AD
STK11	serine/threonine kinase 11	Peutz Jeghers Syndrome, including increased risk for cancers of the gastrointestinal tract, pancreas, cervix, ovary, and breast	AD
TP53	tumor protein p53	Li Fraumeni Syndrome	AD
MUTYH	mutY DNA glycosylase	MUTYH-associated polyposis	AR
NTHL1	nth like DNA glycosylase 1	NTHL1 Tumor Syndrome; NTHL1-Associated Polyposis	AR
POLD1	polymerase (DNA directed), delta 1, catalytic subunit	POLD1 & POLE associated Colorectal Adenomas and Cancer	AD
POLE	polymerase (DNA directed), epsilon, catalytic subunit	POLD1 & POLE associated Colorectal Adenomas and Cancer	AD

^{*}HGNC symbol is the unique identifier assigned to every known human gene by the Human Genome Organization Gene Nomenclature Committee (HGNC).

**Inheritance patterns are autosomal dominant (AD) and/or autosomal recessive (AR).

Table 2: Demographic and clinical characteristics of childhood cancer survivors in CCSS and St. Jude Lifetime Cohort

CRC (n=99)	Total (n=)
	NA

Length of time from primary	
cancer diagnosis to CRC	
diagnosis	
<5 years	NA
5-10 years	NA
11-20 years	NA
21-30 years	NA
30+ years	NA
Alkylating agents, in CED*	
(mg/m ²)	
None	
>0 to < 4,000	
≥ 4,000 to < 8,000	
≥ 8,000	
Procarbazine dose (mg/m²)	
None	
>0 to ≤ 4200	
>4200 to ≤ 7036	
>7036	
Cisplatin (mg/m²)	
None	
1-400	
401-750	
>750	
Abdominal radiotherapy for	
primary malignancy (Gy)	
None	
Yes	
<10	
>10 to <20	
≥ 20	
Pelvic radiotherapy for primary	
malignancy (Gy)	
None	
Yes	
<10	
>10 to < 20	
≥ 20	
Total bady immediation (C)	
Total body irradiation (Gy)	

None	
Yes	
<10	
>10 to < 20	
≥ 20	
Body Mass Index (BMI) at time	
of survey	
Underweight (BMI<18.5)	
Normal weight (18.5≤BMI<25)	
Overweight (25≤BMI<30)	
Obese (BMI≥30)	
Tobacco exposure	
Yes	
No	

^{*}CED=cyclophosphamide-equivalent dose

Table 3: ACMG genotypes

Table 3. ACI	viG genotypes			
		Survivors with CRC	Survivors without	Cananal
		SMN carrying	CRC SMN carrying	General
		variant, n(%)	variants, n(%)	Population, n(%)
Total, n				
Number who				
carry a rare				
variant				
Gene	Variant class			
APC	P/LP			
	LOF			
	Missense			
BMPR1A	P/LP			
	LOF			
	Missense			
Other genes				
listed from Table				
1, etc.*				

^{*} Will list all cancer predisposition genes from the ACMG Secondary Findings, SJCPG₆₀, and 96 genes that are known to be associated with CRC-related cancer predisposition syndromes

Table 4: ACMG genotypes among CRC SMN cases by treatment exposure (n=99)

	- <u>J</u>	Treated with	Treated with	No exposures to
				•
		abdominal/pelvic	procarbazine or	abdominal/pelvic
		radiation and no	cisplatin and no	radiation,
		procarbazine or	abdominal/pelvic	procarbazine, or
Gene	Variant class	cisplatin, n(%)	radiation, n(%)	cisplatin, n(%)
Total, n				
APC	P/LP			
	LOF			
	Missense			
BMPR1A	P/LP			
	LOF			
	Missense			
Other genes listed				
in Table 1, etc.*				

^{*} Will list all cancer predisposition genes from the ACMG Secondary Findings, SJCPG₆₀, and 96 genes that are known to be associated with CRC-related cancer predisposition syndromes

Table 5: Hazard ratios for genetic variants, clinical factors, and treatment exposures for CRC SMN development in childhood cancer survivors

Factors	Hazard ratio	95% confidence interval	p value
Genetic Variant			
P/LP			
Missense			
LOF			
Sex			
Male			
Female			
Age at time of childhood			
cancer diagnosis			
0-4 years old			
5-9 years old			
10-14 years old			
15-20 years old			

Family biotomy of		I
Family history of		
colorectal cancer Yes		
No		
Treatment exposures		
Alkylating agents, in		
CED* (mg/m ²)		
None		
>0 to < 4,000		
≥ 4,000 to < 8,000		
≥ 8,000		
-,		
Procarbazine dose		
(mg/m ²)		
None		
>0 to ≤ 4200		
>4200 to ≤ 7036		
>7036		
Cisplatin (mg/m²)		
None		
1-400		
401-750		
>750		
. 00		
Abdominal radiotherapy		
for primary malignancy		
(Gy)		
None		
Yes		
<10		
>10 to <20		
≥ 20		
Pelvic radiotherapy for		
primary malignancy (Gy)		
None		
Yes		
<10		
>10 to < 20		
≥ 20		

Total body irradiation		
(Gy)		
None		
Yes		
<10		
>10 to < 20		
≥ 20		

Table 6: Self-reported genetic conditions related to CRC and receipt of colonoscopy

Table 6: Self-reported genetic conditions related to CRC and rece	eipt of colonosco
· •	Total, n (%)
Total number of participants who responded to survey, n	
Genetic conditions	
Bloom's syndrome	
Yes	
No	
Not sure	
Familial adenomatous polyposis (FAP or Gardner syndrome)	
Yes	
No	
Not sure	
Other**	
Yes	
No	
Not sure	
Total accept on of a outining outs color and add become a CDC	
Total number of participants who reported having a CRC-	
related genetic condition, n Never received a colonoscopy	
Received a colonoscopy	
Less than 1 year ago	
1-2 years ago	
>2 years to <5 years ago	
5 or more years	+
I had one but I don't recall	+
I don't know if I ever had one or don't know	+
. do. t Allow in 1 ovor had one of don't know	
	_1

^{**}Other includes written responses of _____. We will only include if the condition(s) written is related to CRC (i.e., Lynch syndrome/Hereditary Nonpolyposis Colorectal Cancer, Li-Fraumeni syndrome, etc.)

Figure 1: Cumulative Incidence of Colorectal Cancer Stratified by P/LP Variant Carrier Status in Childhood Cancer Survivors

- X-Axis: Age (in years)
- Y-Axis: Cumulative incidence of CRC (expressed as a proportion).
- Stratification: Separate curves for P/LP variant carriers versus non-carriers.

Figure 2: Forest plots for adjusted hazard ratios and 95% confidence intervals for Colorectal Cancer Risk Factors in Childhood Cancer Survivors

- X-axis: Hazard ratio on a logarithmic scale, with a vertical line at HR = 1 indicating no association.
- Y-Axis: Would list the predictor variables, including:
 - P/LP variant carrier status
 - o Sex
 - Age at childhood cancer diagnosis
 - Family history of CRC
 - Treatment exposures (e.g., radiation, chemotherapy)

REFERENCES:

- 1. Friedman DL, Whitton J, Leisenring W, et al. Subsequent neoplasms in 5-year survivors of childhood cancer: the Childhood Cancer Survivor Study. *J Natl Cancer Inst*. Jul 21 2010;102(14):1083-95. doi:10.1093/jnci/djq238
- 2. Nottage K, McFarlane J, Krasin MJ, Li C, Srivastava D, Robison LL, Hudson MM. Secondary colorectal carcinoma after childhood cancer. *J Clin Oncol*. Jul 10 2012;30(20):2552-8. doi:10.1200/JCO.2011.37.8760
- 3. Henderson TO, Oeffinger KC, Whitton J, et al. Secondary gastrointestinal cancer in childhood cancer survivors: a cohort study. *Ann Intern Med.* Jun 5 2012;156(11):757-66, W-260. doi:10.7326/0003-4819-156-11-201206050-00002
- 4. Reulen RC, Wong KF, Bright CJ, et al. Risk of digestive cancers in a cohort of 69 460 five-year survivors of childhood cancer in Europe: the PanCareSurFup study. *Gut*. Nov 2 2020;doi:10.1136/gutjnl-2020-322237
- 5. Reulen RC, Frobisher C, Winter DL, et al. Long-term risks of subsequent primary neoplasms among survivors of childhood cancer. *JAMA*. Jun 8 2011;305(22):2311-9. doi:10.1001/jama.2011.747
- 6. Bhatia S, Yasui Y, Robison LL, et al. High risk of subsequent neoplasms continues with extended follow-up of childhood Hodgkin's disease: report from the Late Effects Study Group. *J Clin Oncol*. Dec 1 2003;21(23):4386-94. doi:10.1200/JCO.2003.11.059
- 7. Mauri G, Sartore-Bianchi A, Russo AG, Marsoni S, Bardelli A, Siena S. Early-onset colorectal cancer in young individuals. *Mol Oncol*. Feb 2019;13(2):109-131. doi:10.1002/1878-0261.12417
- 8. Biller LH, Ukaegbu C, Dhingra TG, et al. A Multi-Institutional Cohort of Therapy-Associated Polyposis in Childhood and Young Adulthood Cancer Survivors. *Cancer Prev Res (Phila)*. Mar 2020;13(3):291-298. doi:10.1158/1940-6207.CAPR-19-0416
- 9. Peters U, Bien S, Zubair N. Genetic architecture of colorectal cancer. *Gut*. Oct 2015;64(10):1623-36. doi:10.1136/gutjnl-2013-306705

- 10. Allodji RS, Haddy N, Vu-Bezin G, et al. Risk of subsequent colorectal cancers after a solid tumor in childhood: Effects of radiation therapy and chemotherapy. *Pediatr Blood Cancer*. Feb 2019;66(2):e27495. doi:10.1002/pbc.27495
- 11. Teepen JC, Kok JL, van Leeuwen FE, et al. Colorectal Adenomas and Cancers After Childhood Cancer Treatment: A DCOG-LATER Record Linkage Study. *J Natl Cancer Inst*. Jul 1 2018;110(7):758-767. doi:10.1093/jnci/djx266
- 12. Tukenova M, Diallo I, Anderson H, et al. Second malignant neoplasms in digestive organs after childhood cancer: a cohort-nested case-control study. *Int J Radiat Oncol Biol Phys*. Mar 1 2012;82(3):e383-90. doi:10.1016/j.ijrobp.2011.05.069
- 13. Kumar K, Kumar S, Datta K, Fornace AJ, Jr., Suman S. High-LET-Radiation-Induced Persistent DNA Damage Response Signaling and Gastrointestinal Cancer Development. *Curr Oncol.* Jun 7 2023;30(6):5497-5514. doi:10.3390/curroncol30060416
- 14. Kumar S, Suman S, Angdisen J, Moon BH, Kallakury BVS, Datta K, Fornace AJ, Jr. Effects of High-Linear-Energy-Transfer Heavy Ion Radiation on Intestinal Stem Cells: Implications for Gut Health and Tumorigenesis. *Cancers (Basel)*. Oct 4 2024;16(19)doi:10.3390/cancers16193392
- 15. Kim SB, Bozeman RG, Kaisani A, et al. Radiation promotes colorectal cancer initiation and progression by inducing senescence-associated inflammatory responses. *Oncogene*. Jun 30 2016;35(26):3365-75. doi:10.1038/onc.2015.395
- 16. Bai P, Fan T, Sun G, Wang X, Zhao L, Zhong R. The dual role of DNA repair protein MGMT in cancer prevention and treatment. *DNA Repair (Amst)*. Mar 2023;123:103449. doi:10.1016/j.dnarep.2023.103449
- 17. Kaina B, Fritz G, Ochs K, et al. Transgenic systems in studies on genotoxicity of alkylating agents: critical lesions, thresholds and defense mechanisms. *Mutat Res*. Sep 20 1998;405(2):179-91. doi:10.1016/s0027-5107(98)00135-3
- 18. Pearlman R, Frankel WL, Swanson B, et al. Prevalence and Spectrum of Germline Cancer Susceptibility Gene Mutations Among Patients With Early-Onset Colorectal Cancer. *JAMA Oncol*. Apr 1 2017;3(4):464-471. doi:10.1001/jamaoncol.2016.5194
- 19. Djursby M, Madsen MB, Frederiksen JH, et al. New Pathogenic Germline Variants in Very Early Onset and Familial Colorectal Cancer Patients. *Front Genet*. 2020;11:566266. doi:10.3389/fgene.2020.566266
- 20. Frostberg E, Petersen AH, Bojesen A, Rahr HB, Lindebjerg J, Ronlund K. The Prevalence of Pathogenic or Likely Pathogenic Germline Variants in a Nationwide Cohort of Young Colorectal Cancer Patients Using a Panel of 18 Genes Associated with Colorectal Cancer. *Cancers (Basel)*. Oct 12 2021;13(20)doi:10.3390/cancers13205094
- 21. Mao R, Krautscheid P, Graham RP, et al. Genetic testing for inherited colorectal cancer and polyposis, 2021 revision: a technical standard of the American College of Medical Genetics and Genomics (ACMG). *Genet Med.* Oct 2021;23(10):1807-1817. doi:10.1038/s41436-021-01207-9
- 22. Wang Z, Wilson CL, Easton J, et al. Genetic Risk for Subsequent Neoplasms Among Long-Term Survivors of Childhood Cancer. *J Clin Oncol*. Jul 10 2018;36(20):2078-2087. doi:10.1200/JCO.2018.77.8589
- 23. Early-Onset Colorectal Cancer Think Tank. National Cancer Institute. Accessed Jun 12, 2025. https://www.cancer.gov/about-nci/organization/dcb/news/early-onset-crc

- 24. Bomba L, Walter K, Guo Q, et al. Whole-exome sequencing identifies rare genetic variants associated with human plasma metabolites. *Am J Hum Genet*. Jun 2 2022;109(6):1038-1054. doi:10.1016/j.ajhg.2022.04.009
- 25. Genetics of Colorectal Cancer (PDQ(R)): Health Professional Version. *PDQ Cancer Information Summaries*. 2002.
- 26. Yurgelun MB, Kulke MH, Fuchs CS, et al. Cancer Susceptibility Gene Mutations in Individuals With Colorectal Cancer. *J Clin Oncol*. Apr 1 2017;35(10):1086-1095. doi:10.1200/JCO.2016.71.0012
- 27. Valle L, Vilar E, Tavtigian SV, Stoffel EM. Genetic predisposition to colorectal cancer: syndromes, genes, classification of genetic variants and implications for precision medicine. *J Pathol*. Apr 2019;247(5):574-588. doi:10.1002/path.5229
- 28. Landrum MJ, Chitipiralla S, Brown GR, et al. ClinVar: improvements to accessing data. *Nucleic Acids Res.* Jan 8 2020;48(D1):D835-D844. doi:10.1093/nar/gkz972
- 29. McLaren W, Gil L, Hunt SE, et al. The Ensembl Variant Effect Predictor. *Genome Biol*. Jun 6 2016;17(1):122. doi:10.1186/s13059-016-0974-4
- 30. Jurgens SJ, Choi SH, Morrill VN, et al. Analysis of rare genetic variation underlying cardiometabolic diseases and traits among 200,000 individuals in the UK Biobank. *Nat Genet*. Mar 2022;54(3):240-250. doi:10.1038/s41588-021-01011-w
- 31. Cingolani P, Platts A, Wang le L, et al. A program for annotating and predicting the effects of single nucleotide polymorphisms, SnpEff: SNPs in the genome of Drosophila melanogaster strain w1118; iso-2; iso-3. *Fly (Austin)*. Apr-Jun 2012;6(2):80-92. doi:10.4161/fly.19695
- 32. Liu X, Li C, Mou C, Dong Y, Tu Y. dbNSFP v4: a comprehensive database of transcript-specific functional predictions and annotations for human nonsynonymous and splice-site SNVs. *Genome Med.* Dec 2 2020;12(1):103. doi:10.1186/s13073-020-00803-9
- 33. Green DM, Nolan VG, Goodman PJ, et al. The cyclophosphamide equivalent dose as an approach for quantifying alkylating agent exposure: a report from the Childhood Cancer Survivor Study. *Pediatr Blood Cancer*. Jan 2014;61(1):53-67. doi:10.1002/pbc.24679